



# Detection of *Anaplasma* in *Haemaphysalis longicornis* from Hilly Area in Central China

Ke Zhang<sup>1,2</sup>, Xiangdong Jin<sup>1</sup>, Ying Ding<sup>2</sup>, Yujie Chen<sup>2</sup>, Ruixuan Shi<sup>2</sup> and Yuanzhi Wang<sup>3\*</sup>

<sup>1</sup>School of Medical Technology and Engineering, Zhengzhou Railway Vocational and Technical College, Zhengzhou City, 450046, Henan Province, China.

<sup>2</sup>College of Medicine, Pingdingshan University, Pingdingshan City, 467000, Henan Province, China.

<sup>3</sup>College of Medicine, Shihezi University, 832000, Xinjiang, China.

## ABSTRACT

*Anaplasma* are zoonotic intracellular bacteria transmitted mainly by ticks. Ticks and tick-borne *Anaplasma* in the hilly area in Central China are not well studied. During June to July in 2018, ticks were collected from six counties in the hilly area of Funiu Mountains in Central China. All ticks were identified by morphology and confirmed by *16S rDNA* gene sequencing. *Anaplasma* were detected by nested PCR amplification of *16S rDNA* gene. In total, 686 ticks including two species *Haemaphysalis longicornis* (n=683) and *Rhipicephalus microplus* (n=3) were obtained. All of *H. longicornis* tick were clustered into 166 pools (n=1-10), and the minimum infection rate (MIR) of *Anaplasma* was 4.10% (28/683), *Anaplasma phagocytophilum* carriage rate was 3.66% (25/683) and *Anaplasma centrale* was 0.44% (3/683). The *Anaplasma 16S rDNA* gene phylogenetic analysis showed that all *Anaplasma*-positive samples were divided into four clades (Clade 1, 98.23-100% identity to *A. phagocytophilum* from Shandong and Zhejiang Province in China; Clade 2, 98.23% identity to *A. phagocytophilum* from North Korea; Clade 4, 98.48% identity to *A. phagocytophilum* from South Korea; Clade 3, 100% identity to *Anaplasma centrale* from South Korea). In the hilly area of Funiu Mountains in Central China, *H. longicornis* was the dominant tick species with high MIR 3.66% of *A. phagocytophilum* and low MIR 0.44% of *A. centrale*. Ticks and tick-borne *Anaplasma* had high identity to them from other region of Asia. These results indicated that *H. longicornis* tick carried high prevalence *A. phagocytophilum*, which may be a challenge for public health in the study area.

## Article Information

Received 23 April 2023

Revised 05 August 2023

Accepted 24 August 2023

Available online 02 October 2024 (early access)

## Authors' Contribution

KZ and XDJ designed and supervised the experiment and improved the manuscript. HJW, RXS, YJC and YD performed the experiment. YZW reviewed the manuscript and made improvement.

## Key words

*Anaplasma*, *Haemaphysalis longicornis*, Hilly area, Central China

## INTRODUCTION

*Anaplasma* are zoonotic obligate intracellular bacteria mainly transmitted by *Ixodes* spp. ticks, which causes considerable economic losses in the livestock industry and serious public health concerns (Ismail *et al.*, 2010). *Anaplasma* genus includes seven species: *A. phagocytophilum*, *A. ovis*, *A. bovis*, *A. centrale*, *A. marginale*

*A. platys* and *A. capra* (Dumler *et al.*, 2001; Li *et al.*, 2015). *A. phagocytophilum* can cause human granulocytic anaplasmosis with several features ranging from mild illness such as fever, headache, myalgia, malaise, thrombocytopenia, and leukopenia to severe disease such as gastrointestinal, respiratory distress, myocarditis, neurological complications, septic shock-like disease, and even death (Dumler *et al.*, 2007; Bakken and Dumler, 2000). *A. marginale* can cause bovine anaplasmosis with severe anaemia and death in infected cattle and is responsible for economic losses due to high morbidity and mortality, reduced weight gains and milk production, abortions, and treatment costs among cattle worldwide (Battilani *et al.*, 2017). *A. centrale* called *Anaplasma marginale* variety *centrale* before generally causes a milder, less virulent form of the disease. Due to infection with *A. centrale* conferring some cross-protection against *A. marginale*, it has been employed as a live vaccine (Potgieter and Stoltz,

\* Corresponding author: wangyuanzhi621@126.com  
0030-9923/2024/0001-0001 \$ 9.00/0



Copyright 2024 by the authors. Licensee Zoological Society of Pakistan.

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

2004; Theiler, 1912). *A. ovis* can cause anaplasmosis of sheep, goats and wild ruminants, rarely cattle with mild clinical disease (Friedhoff, 1997; Kuttler, 1981; Ryff and Weible, 1964). Although *A. ovis* infects the erythrocyte where it is phenotypically similar to, but does not provide protection against *A. marginale* infection (Splitter, 1956).

Funiu Mountains located in central China is the climate zoning line of the north subtropical and warm temperate zone in China. The mountains in the southeast section usually called hilly area are gradually low and scattered with an altitude of 400-1000 meters, even 200-400 meters, and the slope of the hillside is reduced from 20-40° to 20-35°, even lower than 20°. In the hilly area, traditional sheep, goat and occasionally cattle grazing, and planting of corn, soybean and wheat are both popular. Although ticks and tick-borne *Anaplasma* were reported in North western China (Guo *et al.*, 2016; Yan *et al.*, 2020; Yang *et al.*, 2013), in eastern China (Qin *et al.*, 2018), in southeastern China (Liu *et al.*, 2017), even in Xi'an (Guo *et al.*, 2018), in Hebei province (Zou *et al.*, 2011), in Xinyang in Dabieshan Mountains (Zhuang *et al.*, 2018), in the Funiu Mountains, especially in the hilly area, ticks and tick-borne pathogens are not well studied. The aim of this study was to investigate ticks and tick-borne *Anaplasma* in the hilly area of Funiu Mountains in Central China.

## MATERIALS AND METHODS

### Ticks collection and identification

In 2018, between June and July, at the tick peak time, the ticks were collected from eleven sites of six counties including Wugang, Baofeng, Lushan, Ruzhou, Jiaxian, and Xincheng in the hilly area of Funiu Mountains (Fig. 1). In summer, average daily temperature of the study area ranges from 23°C to 34°C and -2°C to 10°C in winter. The average rainfall is about 790 mm/year. In the study area, the altitude of rolling hills ranges mostly from 200 m to 600 m and the altitude of flatlands is less than 70m.

The parasitic ticks were collected over the livestock's entire body including ears, neck, thorax, armpits, abdomen, interfemur, crissum and so on (Zhang *et al.*, 2016). The free-living ticks were collected by the flagging

method and artificial trapping method (Zhang *et al.*, 2016). All the collected ticks were subjected to starvation for 2-3 days, and a stereomicroscope was used to examine their morphological features including back, abdomen, shield plate, gas door plate, false head base, lateral furrow, and genital orifice. One to ten (mean= 4.06, 686/169) ticks of the same species collected from one site were mixed into one pool with no separation of males and females. Then, the pooled ticks were analyzed individually. Partial *16S rDNA* sequences of the 60 representative ticks, with 1-8 pool of each tick species from each sampling site were sequenced to validate the findings in the morphology of the ticks (Black and Piesman, 1994). A brief summary of this experiment is illustrated in Table I. Then, ticks were stored at -80 °C until DNA extraction.

### DNA extraction

DNA extraction of ticks was carried out following a previously reported procedure (Zhang *et al.*, 2021). In brief, sterile deionized water and 75% ethanol were used for washing and disinfecting of ticks before extraction of DNA. The TIANamp Genomic DNA Kit (TIANGEN Biotech Co., Ltd., Beijing) was used for the extraction of DNA according to the instructions specified by the manufacture.

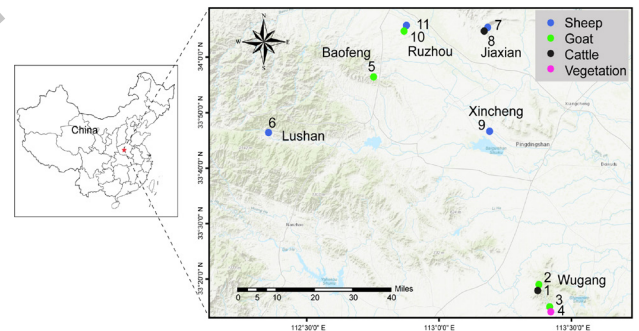


Fig. 1. Map of eleven ticks' collection sites in six counties in the hilly area of Funiu mountains in central China. The purple circle, black circle, green circle, and red circle represent the vegetation, cattle, goat, and sheep, respectively.

Table I. Molecular identification of ticks and tick-borne *Anaplasma* spp.

Ticks and tick-borne agent	Target gene	Primer	Sequences (5'-3')	Tm (°C)	Length (bp)	Ref
Ticks	16S rDNA	<i>16s F</i>	CTGCTCAATGATTTTTTAAATTGCTGTGG	54	460	Black and Piesman (1994)
		<i>16s R</i>	CCGGTCTGAACTCAGATCAAGT			
<i>Anaplasma</i> spp.	16S rDNA	<i>out1</i>	TTGAGAGTTTGATCCTGGCTCAGAACG	55	650	Wen <i>et al.</i> (2002)
		<i>out2</i>	CACCTCTACACTAGGAATTCCGCTATC			
		<i>HGA1</i>	GTCGAACGGATTATTCTTTATAGCTTG	55	390	
		<i>HGA2</i>	TATAGGTACCGTCATTATCTTCCCTAC			

**Table II. Number of tick species recovered from the 11 sampling sites.**

Location	Site No. (Date)	Altitude (m)	Origin	<i>H. longicornis</i> / (♂/♀)	<i>R. microplus</i> / (♂/♀)
Wugang	1#(2018/6/5)	269	Cattle	6(3/3)	2(1/1)
	2#(2018/6/5)	234	Goat	65(29/36)	0(0/0)
	3#(2018/6/5)	221	Goat	92(56/36)	0(0/0)
	4#(2018/6/5)	215	Vegetation	6(2/4)	0(0/0)
Baofeng	5#(2018/7/3)	578	Goat	137(39/98)	0(0/0)
Lushan	6#(2018/7/13)	476	Sheep	110(19/91)	0(0/0)
Jiaxian	7#(2018/7/13)	377	Sheep	153(31/122)	0(0/0)
	8#(2018/7/13)	377	Cattle	25(3/22)	0(0/1)
Xinhua	9#(2018/7/30)	114	Sheep	4(0/4)	0(0/0)
Ruzhou	10#(2018/7/30)	209	Goat	51(17/34)	0(0/0)
	11#(2018/7/30)	202	Sheep	12(0/12)	0(0/0)
Total number				683	3
Percentage (%)				99.56%	0.44%

*Detection of Anaplasma spp. using PCR*

The amplification of the *16S rDNA* gene (390 bp) was carried out via employing nested PCR and sequencing for the molecular identification of *Anaplasma* spp. (Wen *et al.*, 2002). The PCR products of the *16S rDNA* gene for *Anaplasma* was purified using the TIAN gel Midi Purification Kit (TIANGEN, Beijing, China) and then cloned into the pGEM-T Easy vector and sequenced by ABI 3730 with the Sanger sequencing technique on both strands in the Beijing Genomics Institute. A brief summary of these experiments is illustrated in Table I.

*Sequence analysis*

The analysis of gene sequences was carried out with the basic local alignment search tool of NCBI (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). The phylogenetic tree was constructed using Neighbor-Joining approach in the Mega 5.0 software (Tamura *et al.*, 2011; Saitou and Nei, 1987). Bootstrap analysis was tested with 1000 replicates (Felsenstein, 1985).

**RESULTS***Collection and identification of ticks*

A total of 686 ticks were collected from eleven sampling sites in the six counties in the hilly area of Funiu Mountains in Central China (Table II). Of them, 683 (pool=166) ticks were identified as *H. longicornis* and only 3 ticks (pool=3) were identified as *R. microplus* by morphological methods and confirmed by DNA sequencing of the tick *16S rDNA* gene. Compared to the date from GenBank, the *16S rDNA* genes of *H. longicornis* showed 99.51%-100% identity to *H. longicornis* tick from Henan (KJ652225.1), Hubei (KJ710084.1), Beijing (KC203355.1), Hebei (JF979374.1), Gansu (FJ712721.1),

Sichuan (JF979373.1), Shanghai (KP324925.1) from China and Aomori (AB819205.1) from Japan (Fig. 2A). The *16S rDNA* genes of *R. microplus* showed 98.67%-99.76% identity to Henan (KX450285.1) from China and Itanagar (MK621328.1) from Southeast Asia (Fig. 2B).

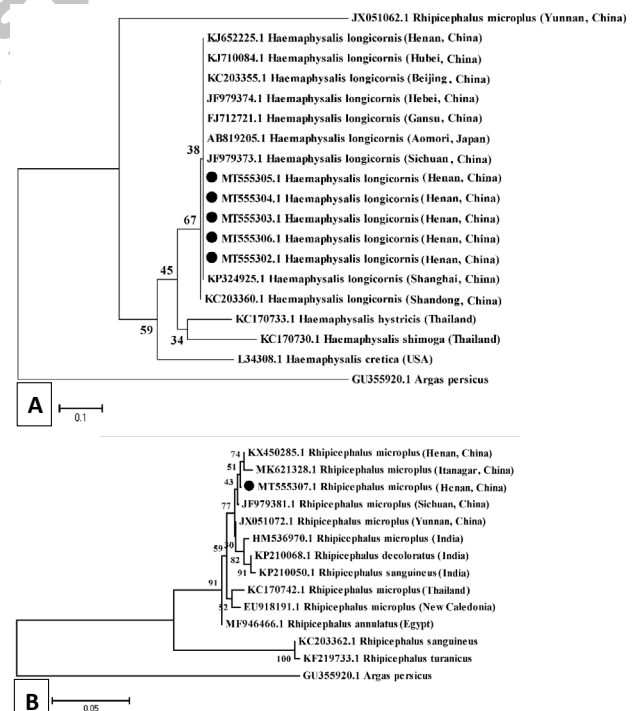


Fig. 2. Phylogenetic analysis of ticks from hilly area in the Funiu Mountains in Central China on the basis of on tick *16S rDNA* (460 bp). The phylogenetic tree for *H. longicornis* (A) and *R. microplus* (B). The bootstrap was 1000 replicates, while at each node, the numbers present bootstrap values. The outgroup is *Argas persicus* (GU355920).





2022; Rehman *et al.*, 2019) and the DNA of *A. centrale* was detected in *D. reticulatus* (Dunaj *et al.*, 2021) and *Ixodes persulcatus* (Wu, 2013). In this study, we detected *A. phagocytophilum* and *A. centrale* in *H. longicornis*.

Of the genus of *Anaplasma*, *A. phagocytophilum* is the agent of human granulocytic anaplasmosis (HGA) (Parola and Raoult, 2001). Domestic animals and wildlife can also be infected by *A. phagocytophilum* (Hartwig *et al.*, 2014; Yang *et al.*, 2013). In China, since the first suspected human case described in 2006 (Zhang *et al.*, 2008), an increasing number of HGA cases have been recorded (Fang *et al.*, 2015). In the present study, high MIR of *A. phagocytophilum* were detected in *H. longicornis* and it is higher than the infection rate of 0.1% from Jiaonan County in Eastern China (Qin *et al.*, 2018) and 2.4% from South Korea (Kim *et al.*, 2003). The diversity could be attributable to different ecological environment of infected ticks or the sensitivity of various primers. Based on our study, high infection rate of *A. phagocytophilum* were in hilly area in Funiu mountains were detected, which was a challenge for public health and the prevalence of *A. phagocytophilum* in herders, livestock and wild life need to be further studied.

*A. centrale* mostly found in wild deer, swine (Portillo *et al.*, 2011; Kawahara *et al.*, 2006), sheep (Zhang *et al.*, 2013) and vectored by several ticks. We detected *A. centrale* in *H. longicornis* from grazing livestock in Funiu Mountains in Central China and marker gene 100% identity to *A. centrale* in *H. longicornis* from South Korea. These results remind us that the people living here have risk to infection with *A. centrale*. Even if *A. centrale* causes a minor infectious disease (Shkap *et al.*, 2002), it still threatens public health especially herdsmen and *A. centrale* cannot be completely ignored.

## CONCLUSION

In the present study, we investigated the ticks and tick-borne *Anaplasma* in the hilly area of Funiu Mountains in central China. *H. longicornis* was the dominant tick species, and the marker genes had a high identity to the tick and tick-borne *Anaplasma* in East Asian-Australasian Flyway of migratory birds. These finding remind us that tick ecology related migration of birds maybe is an important strategy to understand ticks and tick-borne disease and regional cooperatives is needed for prevention and control of ticks and tick-borne disease. In addition, high minimum infection rate of *Anaplasma* especially *A. phagocytophilum* was detected. Our studies suggested ticks and tick borne *Anaplasma* may be a challenge to public health in study area. In future, the prevalence of *A.*

*phagocytophilum* in human and animals need to be further studied.

## DECLARATIONS

### Acknowledgements

We would like to thank the staff of College of Medicine in Shihezi University, Chinese Center for Disease Control and Prevention for their suggestions on improving this study.

### Funding

This work was supported by scientific research projects of Zhengzhou Railway Vocational and Technical College (2104230006, 2402230002).

### IRB approval

This research was conducted with the approval of the Institutional Review Board (IRB). The IRB ensured that all procedures and protocols adhered to ethical guidelines and protected the rights and welfare of research participants. Their oversight was crucial in maintaining the integrity and validity of the study.

### Ethical statement

The study was proved by the Animal Ethics Committee of Zhengzhou Railway Vocational and Technical College (numbered TEMAEC-2018-001).

### Data availability

The nucleotide sequences were submitted to Genbank: *H. longicornis* 16S rDNA gene MT555302-MT555306, *R. microplius* 16S rDNA gene MT555307, *Anaplasma* 16S rDNA gene OQ326846-OQ326850 and *Rickettsiaceae* bacterium 16S rDNA gene OQ383347-OQ383349.

### Statement of conflict of interest

The authors have declared no conflict of interest.

## REFERENCES

- Bakken, J.S. and Dumler, J.S., 2000. Human granulocytic ehrlichiosis. *Clin. Infect. Dis.*, **31**: 554–560. <https://doi.org/10.1086/313948>
- Baldrige, G.D., Scoles, G.A., Burkhardt, N.Y., Schloeder, B., Kurtti, T.J. and Munderloh, U.G., 2009. Transovarial transmission of Francisella-like endosymbionts and *Anaplasma phagocytophilum* variants in *Dermacentor albipictus* (Acari: Ixodidae). *J. Med. Entomol.*, **46**: 625–632. <https://doi.org/10.1603/033.046.0330>
- Battilani, M., De Arcangeli, S., Balboni, A. and

- Dondi, F., 2017. Genetic diversity and molecular epidemiology of *Anaplasma*. *Infect. Genet. Evol.*, **49**, 195–211. <https://doi.org/10.1016/j.meegid.2017.01.021>
- Black, W.C., and Piesman, J., 1994. Phylogeny of hard and soft-tick taxa (Acari: Ixodidae) based on mitochondrial 16S rRNA sequences. *Proc. natl. Acad. Sci.*, **91**: 10034–10038. <https://doi.org/10.1073/pnas.91.21.10034>
- Cao, W.C., Zhao, Q.M., Zhang, P.H., Yang, H., Wu, X.M., Wen, B.H., Zhang, X.T. and Habbema, J.D., 2003. Prevalence of *Anaplasma phagocytophila* and *Borrelia burgdorferi* in *Ixodes persulcatus* ticks from northeastern China. *Am. J. trop. Med. Hyg.*, **68**: 547–550. <https://doi.org/10.4269/ajtmh.2003.68.547>
- Dumler, J.S., Barbet, A.F., Bekker, C.P., Dasch, G.A., Palmer, G.H., Ray, S.C., Rikihisa, Y. and Rurangirwa, F.R., 2001. Reorganization of genera in the families Rickettsiaceae and Anaplasmataceae in the order Rickettsiales: Unification of some species of *Ehrlichia* with *Anaplasma*, *Cowdria* with *Ehrlichia* and *Ehrlichia* with *Neorickettsia*, descriptions of six new species combinations and designation of *Ehrlichia equi* and ‘HGE agent’ as subjective synonyms of *Ehrlichia phagocytophilum*. *Int. J. Syst. Evol. Microbiol.*, **51**: 2145–2165. <https://doi.org/10.1099/00207713-51-6-2145>
- Dumler, J.S., Madigan, J.E., Pusterla, N. and Bakken, J.S., 2007. Ehrlichioses in humans: Epidemiology, clinical presentation, diagnosis, and treatment. *Clin. Infect. Dis.*, **45** (Suppl 1): S45–S51. <https://doi.org/10.1086/518146>
- Dunaj, J., Drewnowska, J., Moniuszko-Malinowska, A., Swiecicka, I. and Pancewicz, S., 2021. First metagenomic report of *Borrelia americana* and *Borrelia carolinensis* in Poland. A preliminary study. *Annls Agric. Environ. Med.*, **28**: 49–55. <https://doi.org/10.26444/aaem/118134>
- Egizi, A., Bulaga-Seraphin, L., Alt, E., Bajwa, W.I., Bernick, J., Bickerton, M., Campbell, S.R., Connally, N., Doi, K., Falco, R.C., Gaines, D.N., Greay, T.L., Harper, V.L., Heath, A.C.G., Jiang, J., Klein, T.A., Maestas, L., Mather, T.N., Occi, J.L., Oskam, C.L., Pendleton, J., Teator, M., Thompson, A.T., Tufts, D.M., Umemiya-Shirafuji, R., VanAcker, M.C., Yabsley, M.J. and Fonseca, D.M., 2020. First glimpse into the origin and spread of the Asian longhorned tick, *Haemaphysalis longicornis*, in the United States. *Zoon. Publ. Hlth.*, **67**: 637–650. <https://doi.org/10.1111/zph.12743>
- Fang, L.Q., Liu, K., Li, X.L., Liang, S., Yang, Y., Yao, H.W., Sun, R.X., Sun, Y., Chen, W.J., Zuo, S.Q., Ma, M.J., Li, H., Jiang, J.F., Liu, W., Yang, X.F., Gray, G.C., Krause, P.J. and Cao, W.C., 2015. Emerging tick-borne infections in mainland China: An increasing public health threat. *Lancet Infect. Dis.*, **15**: 1467–1479. [https://doi.org/10.1016/S1473-3099\(15\)00177-2](https://doi.org/10.1016/S1473-3099(15)00177-2)
- Felsenstein, J., 1985. Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, **39**: 783–791. <https://doi.org/10.1111/j.1558-5646.1985.tb00420.x>
- Friedhoff, K.T., 1997. Tick-borne diseases of sheep and goats caused by *Babesia*, *Theileria* or *Anaplasma* spp. *Parassitologia*, **39**: 99–109.
- Guo, L.P., Jiang, S.H., Liu, D., Wang, S.W., Chen, C.F. and Wang, Y.Z., 2016. Emerging spotted fever group rickettsiae in ticks, northwestern China. *Ticks Tick Borne Dis.*, **7**: 1146–1150. <https://doi.org/10.1016/j.ttbdis.2016.08.006>
- Guo, W.P., Huang, B., Zhao, Q., Xu, G., Liu, B., Wang, Y.H. and Zhou, E.M., 2018. Human-pathogenic *Anaplasma* spp., and *Rickettsia* spp. in animals in Xi’an, China. *PLoS Negl. Trop. Dis.*, **12**: e0006916. <https://doi.org/10.1371/journal.pntd.0006916>
- Hartwig, V., Loewenich, F.D., Schulze, C., Straubinger, R.K., Dausgies, A. and Dyachenko, V., 2014. Detection of *Anaplasma phagocytophilum* in red foxes (*Vulpes vulpes*) and raccoon dogs (*Nyctereutes procyonoides*) from Brandenburg, Germany. *Ticks Tick Borne Dis.*, **5**: 277–280. <https://doi.org/10.1016/j.ttbdis.2013.11.001>
- Hoogstraal, H., Roberts, F.H., Kohls, G.M. and Tipton, V.J., 1968. Review of *Haemaphysalis (Kaiseriana) longicornis* Neumann (Resurrected) of Australia, New Zealand, New Caledonia, Fiji, Japan, Korea, and Northeastern China and USSR, and its parthenogenetic and bisexual populations (Ixodoidea, Ixodidae). *J. Parasitol.*, **154**: 1197–213. <https://doi.org/10.2307/3276992>
- Ismail, N., Bloch, K.C. and McBride, J.W., 2010. Human ehrlichiosis and anaplasmosis. *Clin. Lab. Med.*, **30**: 261–292. <https://doi.org/10.1016/j.cll.2009.10.004>
- Jiang, J.F., Jiang, B.G., Yu, J.H., Zhang, W.Y., Gao, H.W., Zhan, L., Sun, Y., Zhang, X.A., Zhang, P.H., Liu, W., Wu, X.M., Xu, R.M. and Cao, W.C., 2011. *Anaplasma phagocytophilum* infection in ticks, China–Russia border. *Emerg. Infect. Dis.*, **17**: 932–934. <https://doi.org/10.3201/eid1705.101630>
- Kasahara, S., Morimoto, G., Kitamura, W., Imanishi, S. and Azuma, N., 2020. Rice fields along the East Asian–Australasian flyway are important habitats

- for an inland wader's migration. *Sci. Rep.*, **10**: 4118. <https://doi.org/10.1038/s41598-020-60141-z>
- Kawahara, M., Rikihisa, Y., Lin, Q., Isogai, E., Tahara, K., Itagaki, A., Hiramitsu, Y. and Tajima, T.J.A., 2006. Novel genetic variants of *Anaplasma phagocytophilum*, *Anaplasma bovis*, *Anaplasma centrale*, and a novel *ehrlichia* sp. in wild deer and ticks on two major islands in Japan. *Appl. Environ. Microbiol.*, **72**: 1102-1109. <https://doi.org/10.1128/AEM.72.2.1102-1109.2006>
- Keirans, J.E. and Durden, L.A., 2001. Invasion: Exotic ticks (Acari: Argasidae, Ixodidae) imported into the United States. A review and new records. *J. med. Ent.*, **38**: 850-861. <https://doi.org/10.1603/0022-2585-38.6.850>
- Kim, C.M., Kim, M.S., Park, M.S., Park, J.H. and Chae, J.S., 2003. Identification of *Ehrlichia chaffeensis*, *Anaplasma phagocytophilum*, and *A. bovis* in *Haemaphysalis longicornis* and *Ixodes persulcatus* ticks from Korea. *Vector Borne Zoon. Dis.*, **3**: 17-26. <https://doi.org/10.1089/153036603765627424>
- Kuttler, K.L., 1981. Infection of splenectomized calves with *Anaplasma ovis*. *Am. J. Vet. Res.*, **42**: 2094-2096
- Li, H., Zheng, Y.C., Ma, L., Jia, N., Jiang, B.G., Jiang, R.R., Huo, Q.B., Wang, Y.W., Liu, H.B., Chu, Y.L., Song, Y.D., Yao, N.N., Sun, T., Zeng, F.Y., Dumler, J.S., Jiang, J.F. and Cao, W.C., 2015. Human infection with a novel tick-borne *Anaplasma* species in China: A surveillance study. *Lancet Infect Dis.*, **15**: 663-670. [https://doi.org/10.1016/S1473-3099\(15\)70051-4](https://doi.org/10.1016/S1473-3099(15)70051-4)
- Liu, X.Y., Gong, X.Y., Zheng, C., Song, Q.Y., Chen, T., Wang, J., Zheng, J., Deng, H.K. and Zheng, K.Y., 2017. Molecular epidemiological survey of bacterial and parasitic pathogens in hard ticks from eastern China. *Acta Trop.*, **167**: 26-30. <https://doi.org/10.1016/j.actatropica.2016.12.010>
- Ngnindji-Youdje, Y., Diarra, A.Z., Lontsi-Demano, M., Tchuinkam, T., Parola, P., 2022. Detection of tick-borne pathogens in ticks from cattle in western highlands of cameroon. *Microorganisms*, **10**: 1957. <https://doi.org/10.3390/microorganisms10101957>
- Parola, P. and Raoult, D., 2001. Ticks and tick-borne bacterial diseases in humans: An emerging infectious threat. *Clin. Infect. Dis.*, **32**: 897-928. <https://doi.org/10.1086/319347>
- Portillo, A., Pérez-Martínez, L., Santibáez, S., Santibáez, P., Palomar, A.M. and Oteo, J.A., 2011. *Anaplasma* spp. in wild mammals and *Ixodes ricinus* from the North of Spain. *Vector Borne Zoon. Dis.*, **11**: 3-8. <https://doi.org/10.1089/vbz.2009.0214>
- Potgieter, F.T. and Stoltsz, W.H., 2004. *Infectious diseases of livestock*. Oxford University Press, Cape Town, South Africa. pp. 594-616.
- Qin, X.R., Han, F.J., Luo, L.M., Zhao, F.M., Han, H.J., Zhang, Z.T., Liu, J.W., Xue, Z.F., Liu, M.M., Ma, D.Q., Huang, Y.T., Sun, Y., Sun, X.F., Li, W.Q., Zhao, L., Hao, Y. and Yu, X.J., 2018. *Anaplasma* species detected in *Haemaphysalis longicornis* tick from China. *Ticks Tick Borne Dis.*, **9**: 840-843. <https://doi.org/10.1016/j.ttbdis.2018.03.014>
- Rehman, A., Conraths, F.J., Sauter-Louis, C., Krücken, J. and Nijhof, A.M., 2019. Epidemiology of tick-borne pathogens in the semi-arid and the arid agro-ecological zones of Punjab province, Pakistan. *Transbound. Emerg. Dis.*, **66**: 526-536. <https://doi.org/10.1111/tbed.13059>
- Ryff, J.F. and Weibel, J.L., 1964. Thomas GM. Relationship of ovine to bovine Anaplasmosis. *Cornell Vet.*, **54**: 407-414.
- Saitou, N. and Nei, M., 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.*, **4**: 406-425.
- Shkap, V., Molad, T., Fish, L. and Palmer, G.H., 2002. Detection of the *Anaplasma centrale* vaccine strain and specific differentiation from *Anaplasma marginale* vaccinated and infected cattle. *Parasitol. Res.*, **88**: 546-552. <https://doi.org/10.1007/s00436-002-0612-9>
- Somveille, M., Manica, A., Butchart, S.H. and Rodrigues, A.S., 2013. Mapping global diversity patterns for migratory birds. *PLoS One*, **8**: e70907. <https://doi.org/10.1371/journal.pone.0070907>
- Splitter, E.J., Anthony, H.D., Twiehaus, M.J., 1956. *Anaplasma ovis* in the United States; experimental studies with sheep and goats. *Am. J. Vet. Res.*, **17**: 487-491.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., Kumar, S., 2011. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.*, **28**: 2731-2739. <https://doi.org/10.1093/molbev/msr121>
- Theiler, A., 1912. Gallsickness of imported cattle and the protective inoculation against this disease. *Agric. J. Union S. Africa.*, **3**: 1-11.
- Wen, B., Jian, R., Zhang, Y. and Chen, R., 2002. Simultaneous detection of *Anaplasma marginale* and a new *Ehrlichia* species closely related to *Ehrlichia chaffeensis* by sequence analyses of 16S Ribosomal DNA in *Boophilus microplus* ticks from tibet. *J. clin. Microbiol.*, **40**: 3286-3290. <https://doi.org/10.1128/JCM.40.9.3286-3290.2002>

- Wu, R.T., 2013. Parsing and identification of rickettsia of eastern wild deer and ticks insects. *J. Inner Mongol. Agric. Univ.*, **34**: 1-6.
- Yan, Y.Q., Jiang, Y.X., Tao, D.Y., Zhao, A.Y., Qi, M. and Ning, C.S., 2020. Molecular detection of *Anaplasma* spp. in dairy cattle in southern Xinjiang, China. *Vet. Parasitol. Reg. Stud. Reports.*, **20**: 100406. <https://doi.org/10.1016/j.vprsr.2020.100406>
- Yang, J.F., Liu, Z.J., Guan, G.Q., Liu, Q., Li, Y.Q., Chen, Z., Ma, M.L., Liu, A.H., Ren, Q.Y., Luo J.X., and Yin, H., 2013. Prevalence of *Anaplasma phagocytophilum* in ruminants, rodents and ticks in Gansu, northwestern China. *J. med. Microbiol.*, **62**: 254-258. <https://doi.org/10.1099/jmm.0.046771-0>
- Zhang, G., Sun, X., Zhao, Y., Liu, X., Zheng, Z., Sun, Y. and Liu, R., 2013. Prevalence of *Anaplasma* spp. infection in a desert landscape region of Heshuo, Xinjiang. *Chin. J. Epidemiol.*, **34**: 147-151.
- Zhang, K., Li, A.J., Wang, Y., Zhang, J.J., Chen, Y.J., Wang, H.J., Shi, R.X. and Qiu, Y.H., 2021. Investigation of the presence of *Ochrobactrum* spp. and *Brucella* spp. in *Haemaphysalis longicornis*. *Ticks Tick Borne Dis.*, **12**: 101588. <https://doi.org/10.1016/j.ttbdis.2020.101588>
- Zhang, L., Liu, Y., Ni, D., Li, Q., Yu, Y., Yu, X.J., Wan, K., Li, D., Liang, G., Jiang, X., Jing, H., Run, J., Luan, M., Fu, X., Zhang, J., Yang, W., Wang, Y., Dumler, J.S., Feng, Z., Ren, J. and Xu, J., 2008. Nosocomial transmission of human granulocytic anaplasmosis in China. *J. Am. med. Assoc.*, **300**: 2263–2270. <https://doi.org/10.1001/jama.2008.626>
- Zhang, Y.Z. and Xu, J.G., 2016. The emergence and cross species transmission of newly discovered tick-borne Bunyavirus in China. *Curr. Opin. Virol.*, **16**: 126–131. <https://doi.org/10.1016/j.coviro.2016.02.006>
- Zhuang, L., Du, J., Cui, X.M., Li, H., Tang, F., Zhang, P.H., Hu, J.G., Tong, Y.G., Feng, Z.C. and Liu, W., 2018. Identification of tick-borne pathogen diversity by metagenomic analysis in *Haemaphysalis longicornis* from Xinyang, China. *Infect. Dis. Poverty*, **7**: 45. <https://doi.org/10.1186/s40249-018-0417-4>
- Zou, Y.X., Jin, H.T., Wang, Q.Y., Fu, Z.X., Gao, H.W., Liu, P.P., Liu, Q. and Chen, L.F., 2011. Molecular detection of *Anaplasma phagocytophilum* in *Ixodid* ticks in Hebei Province, China. *Vector Borne Zoon. Dis.*, **11**: 1323-1327. <https://doi.org/10.1089/vbz.2010.0253>